

Original article

***In Vitro* Antimicrobial Effectiveness of Calcium Hydroxide and Its Combination with Sodium Hypochlorite against Enterococcus Faecalis and Candida Albicans**

Mohamed Issa^{1*}, Najwa Marghani²

¹Department of Conservative Dentistry and Endodontics, Faculty of Dentistry, University of Tripoli, Tripoli, Libya

²Department of Laboratory, Tripoli University Hospital, Tripoli, Libya

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Corresponding Email. elfoghi2004@yahoo.com

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ABSTRACT

The aim of this *in vitro* study was to evaluate and compare the antimicrobial effect of calcium hydroxide ($\text{Ca}(\text{OH})_2$) and its combination with 5% sodium hypochlorite (NaOCL) as intracanal medicaments against *Enterococcus faecalis* (*E. faecalis*) and *Candida albicans* after one, three and seven days. Inoculate of these organisms (*E. faecalis* and *Candida albicans*) were used to make lawn cultures on Mueller-Hinton with 5% defibrinated sheep blood agar and Mueller-Hinton agar. 60 wells with a depth of 4 mm were prepared for *E. faecalis*. Then 20 wells filled with $\text{Ca}(\text{OH})_2$, 20 wells filled with combination of $\text{Ca}(\text{OH})_2$ and 5% NaOCL, and last 20 wells were filled with distilled water as control. A similar procedure was conducted for *Candida albicans*. The zone of inhibition for each material used against a particular organism were measured and recorded after 1, 3, and 7 days. $\text{Ca}(\text{OH})_2$ in combination with 5% NaOCL is more effective than $\text{Ca}(\text{OH})_2$ alone against *E. faecalis* and *Candida albicans* at different time periods. *E. faecalis* was more resistant than *Candida albicans* to intracanal medicaments used. The antimicrobial effect depends on how long it remains inside the root canal. Mixing of $\text{Ca}(\text{OH})_2$ with 5% NaOCL had antimicrobial effect on both *E. faecalis* and *Candida albicans* and it was more effective against *Candida albicans*.

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INTRODUCTION

Microorganisms are the cause of apical inflammatory lesions, and the most significant goal of an endodontic treatment is the maximum reduction or elimination of microorganisms and their products from the root canal system. It is difficult to eliminate all microorganism from an infected root canal system by mechanical instrumentation alone [1]. The known recovery of *E. faecalis* and *Candida albicans* from failed root canals of teeth in which previous treatment has failed is notable [2,3]. Therefore, chemical irrigation and disinfection are necessary to remove microorganisms, their byproducts and debris from root canal. Intracanal medicaments may perform these roles by remaining in the root canal during treatment appointments [4].

$\text{Ca}(\text{OH})_2$ is widely used as an intracanal medication due to its properties such as its tissue dissolving capability, antimicrobial effects, biocompatibility, maintenance in root canal for long time and continuous release of OH-ions [4-7]. However, it has a limited action against some microorganisms, particularly *E. faecalis* [8] and *Candida albicans* [9] and requires 60 days to have antimicrobial effect on *Candida albicans* and *E. faecalis* [10]. Some studies have reported

the failure of calcium effectively as they tolerate high values of pH [11,12]. In vitro studies have shown that when in direct contact, $Ca(OH)_2$ eliminated microorganisms present in the root canals [13].

NaOCL is currently the most commonly used irrigant with excellent tissue dissolving and antibacterial activities [14,15,16,17]. The antimicrobial effectiveness of NaOCL, based in its high pH (hydroxyl ion action). It interferes with the cytoplasmic membrane integrity with an irreversible enzymatic inhibition, biosynthetic alterations in cellular metabolism and phospholipid degradation. NaOCL can dissolve remnant debris in canal, a property which is desired from an intracanal medicament [18]. Valera et al., [19] stated that 1% NaOCL was an effective in immediate reduction of *Candida albicans* and *E. faecalis* counts after root canal preparation. Other studies have confirmed the advantage of NaOCL with other materials. It has been found that the association of NaOCL with $Ca(OH)_2$ shows equal antibacterial activity to $Ca(OH)_2$ and chlorhexidine combination. Zehnder et al., [20] revealed that a quicker antimicrobial activity for $Ca(OH)_2$ with NaOCL in comparison to $Ca(OH)_2$ with water. Farhad et al., [21] stated that the antibacterial activity of $Ca(OH)_2$ and NaOCL did not differ significantly from $Ca(OH)_2$ and chlorhexidine or $Ca(OH)_2$ and water. Suhad et al [22] reported that adding of NaOCL enhance the bactericidal of $Ca(OH)_2$ against microorganisms. In contrast Verrisimo et al [23] concluded that NaOCL showed the worst performance when used alone as intracanal medicament. This is probably occurred because NaOCL losses its antimicrobial properties and becomes ineffective inside the canal after short time. This study was to evaluate and compare the antimicrobial effect of calcium hydroxide ($Ca(OH)_2$) and its combination with 5% sodium hypochlorite (NaOCL) as intracanal medicaments against *Enterococcus faecalis* (*E. faecalis*) and *Candida albicans* after one, three and seven days.

METHODS

In this study, $Ca(OH)_2$ (i-dental, Lithuania), 5% NaOCL (DR. DEPPE, Germany), and microorganisms (*E. faecalis* and *Candida albicans*) were used. Agar plates were prepared and used for microorganisms. Aseptic culture was sub-cultured in sterile nutrient broth and incubated at 37°C for 24 hours. The inoculate of the strains of *E. faecalis* and *Candida albicans* were prepared with distilled water. This inoculum was used to prepare a lawn culture of the organisms using sterile cotton swabs on Mueller-Hinton Agar with 5% defibrinated sheep blood.

A total of 15 plates of Muller-Hinton Agar with 5% defibrinated sheep blood agar for *E. faecalis* were made. Then 4 wells for each plate of diameter 4 mm and depth 4 mm were punched with a sterile punch (60 wells inside 15 plates). The first 20 wells were filled with $Ca(OH)_2$ paste prepared with distilled water (1g $Ca(OH)_2$ powder mixed with 1 ml of distilled water), and the second 20 wells were filled with a combination of 5% NaOCL and $Ca(OH)_2$ paste, and the last 20 wells were filled with distilled water as control. All materials were prepared and used according to the manufacturer's instructions

A total of 15 plates of Mueller-Hinton Agar for *Candida albicans* were made. A similar procedure was conducted to prepare and fill the 60 wells for *Candida albicans* (20 wells filled with $Ca(OH)_2$ prepared with distilled water, 20 wells filled with a combination of 5% NaOCL, and $Ca(OH)_2$ and 20 wells filled with distilled water as control).

All the plates were incubated overnight at 37°C. The specimens were examined after 24 hours, 72 hours and 7 days respectively. The growth inhibition zones around each intracanal medicament were evidenced by the lack of microbial colonization (cleaning of the agar) around each agar well. The zones of inhibition were measured with a transparent ruler, and the 4 mm diameter of the wells was included in the measurement. The wider diameter of the inhibition zone was indicated the higher antimicrobial activity of the intracanal medicament.

The collected data was statistically analyzed by using ANOVA, and the t test. The level of significance was chosen at $P=0.05$. All statistical analyses were carried out with the SPSS 25 software system.

RESULTS

The mean and standard deviation values for the antimicrobial activity (inhibition zone in mm) of $Ca(OH)_2$ and its combination with 5% NaOCL against *E. faecalis* and *Candida albicans* are presented in Table 1 and Figure 1.

The antimicrobial activity of $Ca(OH)_2$ against *E. faecalis* increased by time, and the zones of microbial growth inhibition (in mm) obtained after one day were 18 ± 0.9176 , increased after three days at 20.2 ± 1.1516 , and reached their maximum after seven days at 21.75 ± 1.2513 .

The antimicrobial activity of $Ca(OH)_2$ and 5% NaOCL combination against *E. faecalis* increased with time, and the inhibition zone diameters obtained after one day were 19 ± 0.9176 and increased after three days to 21.4 ± 1.6026 and reached to maximum after seven days were 23.4 ± 1.6026

The antimicrobial efficacy of $Ca(OH)_2$ against *Candida albicans* increased with time, and the diameters of the zones of microbial growth inhibition (in mm) obtained after one day were 20.05 ± 0.8870 and increased after three days to 27.2 ± 1.5761 and reached their maximum after seven days were 30.2 ± 0.8335 .

The antimicrobial efficacy of $Ca(OH)_2$ and NaOCL combinations against *Candida albicans* increased by time, and the diameters of the zones of microbial growth inhibition obtained after one day were 32.85 ± 2.0589 , increased after three days to 35 ± 1.6543 , and reached of their maximum after seven days were 36.8 ± 2.0416 . No antimicrobial activity was observed in the control against for both microorganisms at all time intervals. The adding of 5% NaOCL to $Ca(OH)_2$ recorded higher antimicrobial mean values (inhibition zone) than $Ca(OH)_2$ alone against *E. faecalis* in all three different investigated periods (24 hours, 72 hours, and 7 days). The inhibition zone values increased with time.

There was a statistically significant difference between the inhibition zones of $Ca(OH)_2$ alone and its combination with 5% NaOCL for *E. faecalis* after 24 hours, 72 hours, and 7 days ($P < 0.05$). That is, in all of them, equal to ($P = 0.01$). There was a statistically significant difference between the inhibition zones of all tested materials against *E. faecalis* at all three periods of time ($P < 0.05$) which is equal to ($P = 0.000$). likewise, there was also a statistically significant difference between the control group and each tested material against *E. faecalis* overall time periods ($P < 0.05$), which is equal to ($P = 0.000$).

The antimicrobial activity of 5% NaOCL combined with $Ca(OH)_2$ exhibited the higher inhibition zone against *Candida albicans* than $Ca(OH)_2$ after 24 hours, 72 hours, and 7 days with significant statistical difference ($P < 0.05$). That is, in all of them, equal to ($P = 0.000$). All tested materials showed significant statistical difference in the antimicrobial activities against *Candida albicans* in three different time periods ($P < 0.05$), with P-value of 0.000. The control group revealed significant statistical difference with each tested material in the antimicrobial activity against *Candida albicans* over all the tested time periods ($P < 0.05$), with a P-value of 0.000. All tested materials $Ca(OH)_2$ and its combination with 5% NaOCL were more effective as antimicrobial against *Candida albicans* than *E. faecalis* at 24 hours, 72 hours, and 7 days. In other words, all tested materials have superior antifungal than antibacterial effects over all three periods of time. A statistical analysis showed significant differences in all tested materials between *Candida albicans* and *E. faecalis* at all periods of time ($P < 0.05$), which is equal to ($P = 0.000$).

Table 1. Mean values and standard deviation (SD) of the antimicrobial efficacy of $Ca(OH)_2$ and its combination with NaOCL against *E. faecalis* and *Candida albicans*

Tested Materials	<i>E. faecalis</i>				<i>Candida albicans</i>			
	Days	NO.	Mean	SD	Days	NO.	Mean	SD
$Ca(OH)_2$	1	20	18	0.9176	1	20	20.05	0.8870
	3	20	20.2	1.1516	3	20	27.2	1.5761
	7	20	21.75	1.2513	7	20	30.2	0.8335
$Ca(OH)_2$ combined with NaOCL	1	20	19	0.9176	1	20	32.85	2.0589
	3	20	21.4	1.6026	3	20	35	1.6543
	7	20	23.4	1.6026	7	20	36.8	2.0416

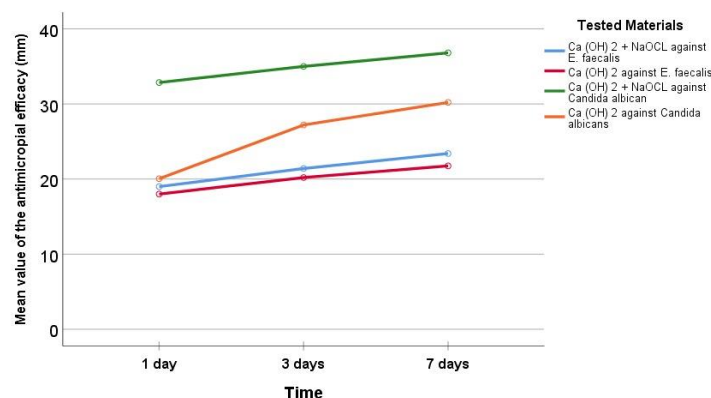


Figure 1. Mean values of the antimicrobial efficacy between *E. faecalis* and *Candida albicans* of tested materials after 1 day, 3 days, and 7 days.

DISCUSSION

Radical elimination of microorganisms from infected root canal is difficult task and various measures have been recommended to reduce the numbers of endodontic microorganisms, including the use of intracanal medicaments and irrigation regimens [24].

$Ca(OH)_2$ has been established as antimicrobial agent and it was reported that it may be the best available inter appointment medication [25]. Its antimicrobial activity is related to the release of hydroxyl ions with consequent pH

increase and the inactivation of enzymes of the microorganisms [26]. NaOCL is antimicrobial agent frequently used in root canal therapy as irrigant as well as intracanal medicament [20]. It was reported that hypochlorite acid is formed in the presence water containing active chlorine, a powerful oxidizing agent that produces an antimicrobial effect by disturbing the metabolic functions of microorganism cells [15]. These materials were investigated in this study.

E. faecalis and *Candida albicans* have been repeatedly identified as the species most commonly recovered from root canals undergoing retreatment in cases of failed endodontic therapy and canals with persistent infections [27,28]. These microorganisms were investigated in this study. The present study resulted that $Ca(OH)_2$ had antimicrobial activity against *E. faecalis* as demonstrated by the formation of growth inhibition zones. The explanation of this could be related to the lethal effects of hydroxyl ions by unabling the bacteria to survive in the highly alkaline environment [29].

Results of the present study are in line with the findings of Delgado et al., [30] who found that $Ca(OH)_2$ has antimicrobial effects on *E. faecalis* and Mehrvarzfar et al., [31] who reported that $Ca(OH)_2$ exhibited antimicrobial effects against *E. faecalis* after 24-, 48-, and 72-hours intervals. On the other hand, the results are in disagreement with the previous studies who concluded that $Ca(OH)_2$ has limited antimicrobial action against *E. faecalis* [8] and requires 60 days to have antimicrobial effect against *E. faecalis* [10]. These inconsistencies might be due to the difference in particle proportion and percentage, the type of $Ca(OH)_2$ tested, and the antimicrobial test used [31].

In the current study, the results of combination of $Ca(OH)_2$ with NaOCL revealed high antimicrobial activity against *E. faecalis*. It also showed that the antimicrobial activity of its combination is greater than $Ca(OH)_2$ by itself with a significant effect on *E. faecalis*. The antimicrobial activity of NaOCL based on its high pH (hydroxyl ion action). It also interferes with the cytoplasmic membrane integrity with an irreversible enzymatic inhibition, biosynthetic alterations in cellular metabolism and phospholipid degradation [18]. Several studies have confirmed the advantage of combination of $Ca(OH)_2$ with NaOCL. Zehnder et al., [20] reported a quicker antimicrobial effect for $Ca(OH)_2$ mixed with NaOCL in comparison to $Ca(OH)_2$ with water. Suhad et al., [22] concluded that when $Ca(OH)_2$ is mixed with NaOCL, the antimicrobial efficacy of the mixture is greater than $Ca(OH)_2$ by itself. Farhad et al., [21] found that high antimicrobial effectiveness of $Ca(OH)_2$ in combination with NaOCL against *E. faecalis*. Valera et al., [19] revealed that effectiveness of NaOCL against *E. faecalis* were noticed. These results are in line with our study.

The present study revealed that $Ca(OH)_2$ showed high antimicrobial activity against *Candida albicans*. The explanation of this could be related to the action of hydroxyl ions on the inactivation of enzymes of the cytoplasmic membrane of microorganisms and causing toxic effects to their cells [26]. The present results were in agreement with the findings of Valera et al., [19] and McHugh et al., [32] who reported that $Ca(OH)_2$ minimize the growth of *Candida albicans*. On the other hand, our findings were disagreed with finding of Attia et al., [33]. They observed that $Ca(OH)_2$ had no antifungal activity, so ineffective in eliminating *Candida albicans*. $Ca(OH)_2$ has a limited antimicrobial action against *Candida albicans* [9]. Furthermore, Estrela et al., [10] finding did not in consistent with our result, when they concluded that $Ca(OH)_2$ requires 60 days to have an antimicrobial effect on *Candida albicans*. The explanation for this could be related to testing methodology and materials properties (Attia et al., [33]).

The current study also showed stronger antimicrobial effect for $Ca(OH)_2$ with NaOCL in comparison to $Ca(OH)_2$ mixed water. The explanation of this could be attributed to the formation of hypochlorite acid which containing active chlorine and a powerful oxidizing agent that produces an antimicrobial effect by irreversible oxidation of hydrosulphuric group of essential enzymes disturbing the metabolic function of microorganism cell [15]. The results of present study were in line with previous studies done by Hulsmann et al., [34] and Valera et al., [19] who found that NaOCL has been proven to be an effective in eliminating *Candida albicans*. However, Happsalo & shen [35] and Abbaszadegan et al., [36] reported that NaOCL has limited effect in eliminating microorganisms, which are in contrary with our study. This result may be attributed to the different *Candida* strains, materials properties and antimicrobial test method.

The present results revealed that all the tested materials ($Ca(OH)_2$ and its combination with NaOCL) record higher antimicrobial activity against *Candida albicans* than *E. faecalis* at all periods of time (1,3, and 7 days). This result could be attributed to the fact that *E. faecalis* was more resistant to high pH environments than another microbe. It can withstand abroad pH range of up to 11.5 and continue to exist after root canal treatment [37]. It is also the bacterium might have other defense mechanisms to withstand this condition and prevent pH damage [38].

CONCLUSION

Both $Ca(OH)_2$ and its combination with 5% NaOCL showed antimicrobial and antifungal effect. *E. faecalis* was more resistant than *Candida albicans* to both tested groups. The combination of $Ca(OH)_2$ with 5% NaOCL has higher antimicrobial activity than $Ca(OH)_2$ mixed with water against both *E. faecalis* and *Candida albicans*. The antimicrobial property was increased over time for all tested materials. The antimicrobial effect depends on how long it remains inside the root canal.

Conflicts of Interest

The authors declare no conflicts of interest.

REFERENCES

1. Bystrom A, Sundqrst G. Bacteriological evaluation of the efficacy of mechanical root canal instrumentation in endodontic therapy. *Scand J Dent Res*. 1981;89(4): 321-328.
2. Sen BH, Piskin B, Demirci T. Observation of bacteria and fungus in infected root canals and dentinal tubules by SEM. *Dent Traumatol*. 1995;11(1): 6-9.
3. Sundqrst G, Figdor D, Persson S, Sjogren U. Microbiologic analysis of teeth with failed endodontic treatment and the outcome of conservative re-treatment. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 1998;85(1): 86-93.
4. Kumar A, Tamanna S, Iftekhhar H. Intra canal medicaments – their use in modern endodontics. *J Oral Res and Review*. 2019;11(2): 94-99.
5. Menezes MM, Valera MC, Jorge AO, Koga-Ito CY, Camargo CH, Mancini MN. In vitro evaluation of the effectiveness of irrigants and intra canal medicaments on microorganisms within root canals. *Int Endod J*. 2004;37(5): 311-319.
6. Athanassiadis B, Abbott PV, Walsh LJ. The use of calcium hydroxide, antibiotics and bioacids as antimicrobial medicaments in endodontics. *Aust Dent J*. 2007;52(2): 64-82.
7. Mustafa M, Saujanya KP, Jain D, Sajjanshetty S, Arun A, Uppin L, Kadri M. Role of calcium hydroxide in endodontics: A review. *Global Journal of Medicine and public health*. 2012;1(1): 66-70.
8. Estrela C, Pimena FC, Ito IY, Bammann LL. Antimicrobial evaluation of calcium hydroxide in infected dentinal tubules. *J Endod*. 1999;25(6): 416-418.
9. Ballal V, Kundabala M, Acharya S, Ballal M. Antimicrobial action of calcium hydroxide, chlorhexidine and their combination on endodontic pathogens. *Aust Dent J*. 2007;52(2): 118-121.
10. Estrela C, Cynthia RA, Pecora JD. A study of the time necessary for calcium hydroxide to eliminate microorganisms in infected canals. *J Appl Oral Sci*. 2003;11(2): 133-137.
11. Gomes BPFA, Ferraz CCR, Rosalen PL. Microbial susceptibility to Ca(OH)₂ pastes and their vehicles. *J Endod*. 2002;28(11): 758-761.
12. Pinheiro ET, Gomes BPFA, Ferraz CCR, Sousa ELR, Teixeira FB, Souza-Filho FJ. Microorganisms from canals of root filled teeth with periapical lesions. *Int Endod J*. 2003;36(1): 1-11.
13. Souza-Filho FJ, Soares AJ, Vianna ME, Zaia AA, Ferraz CC, Gomes BP. Antimicrobial effect and pH of chlorhexidine gel and calcium hydroxide alone and associated with other materials. *Braz Dent J*. 2008;19(1): 29-33.
14. Berutti E, Marini R, Angeretti A. Penetration ability of different irrigants into dentinal tubules. *J Endod*. 1997;23(12): 725-727.
15. Siqueira JFJR, Rocas IN, Favieri A, Lima KC. Chemomechanical reduction of the bacterial population in the root canal after instrumentation and irrigation with 1 %, 2.5 % and 5.25 % sodium hypochlorite. *J Endod*. 2000;26(6): 331-334.
16. Thomas L, Maillard JY, Lambert RT, Russell AD. Development of resistance to chlorhexidine diacetate in pseudomonas aeruginosa and the effect of a residual concentration. *J Hosp Infect* 2000;46(4): 297-303.
17. Valera MC, Rego JM, Jorge AOC. Effect of sodium hypochlorite and five intra canal medications on *Candida albicans* in root canals. *J Endod*. 2001;27(6): 401-408.
18. Estrela C, Estrela CRA, Barbin EL, Spano LCE, Marchesan MA, Pecora JD. Mechanism of action of sodium hypochlorite. *Braz Dent J*. 2002;13(2): 113-117.
19. Valera MC, Siva KC, Maekawa LE, Carvalho CA, Koga-Ito CY, Camargo CH. Antimicrobial activity of sodium hypochlorite associated with intra canal medication for *Candida albicans* and *Enterococcus faecalis* inoculated in root canals. *J Appl Oral Sci*. 2009;17(6): 555-559.
20. Zehnder MI, Grawehr V, Hasselgren G, Waltimo T. Tissue-dissolution capacity and dentin-disinfecting potential of calcium hydroxide mixed with irrigating solutions. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 2003;96(5): 608-613.
21. Farhad AR, Barekatin B, Allameh M, Narimani T. Evaluation of the antibacterial effect of calcium hydroxide in combination with three different vehicles. An invitro study. *Dent Res J*. 2012;9(2): 167-172.
22. Suhad JH, Israa KA, Nibrass TA, Mahdi AA. Antibacterial activity of calcium hydroxide combined with chlorhexidine or sodium hypochlorite against gram positive and gram negative bacteria. *J Natural Sci Res*. 2014;4(12): 55- 61.
23. Verissimo R, Gurgel-Filho E, De-Deus G, Coutinho-Filho T, de Souza-Filho F. Coronal leakage of four intra canal medications after exposure to human saliva in the presence of a temporary filling material. *Indian J Dent Res*. 2010;21(1): 35-39.
24. Gomes BPI, Lilley JD, Dracker BD. Variations in the susceptibility of components of the endodontic microflora to biomechanical procedures. *Int Endod J*. 1996;29(4): 235-241.
25. Law AI, Messer H. An evidence-based analysis of the antibacterial effectiveness of intra canal medicaments. *J Endod*. 2004;30(10): 689-694.
26. Siqueira JFJR, Uzeda M. Desinfection by calcium hydroxide pastes of dentinal tubules infected with two obligate and one facultative anaerobic bacterium. *J Endod*. 1996;22(12): 674-676.

27. Gopikrishna V, Kandaswamy D, Jeyavel RK. Comparative evaluation of the antimicrobial efficacy of five endodontic root canal sealers against *Enterococcus faecalis* and *Candida albicans*. *Journal of Conservative Dentistry*. 2006;9(1): 2-7.
28. Mathew R, Sukumaran, AS, Singh P, Varughese AV. Evaluation of the efficacy of different intra canal medicaments against *Candida albicans* and *Enterococcus faecalis* – An invitro study. *Indian J Dent Res*. 2022;33(4): 440-445.
29. Estrela C, Holland R. Calcium hydroxide: Study based on scientific evidences. *J Applied Oral Sci*. 2003;11: 269-282.
30. Delgado R, Gasparoto T, Sipert C, Pinheiro C, Moraes IG, Garcia RB, et al. Antimicrobial effects of calcium hydroxide and chlorhexidine on *Enterococcus faecalis*. *J Endod*. 2010;36(8): 1389-1393.
31. Mehrvarzfar P, Akhavan H, Rastgarian H, Akhlagi NM, Soleymanpour R, Ahmadi A. An in vitro comparative study on the antimicrobial effects of bioglass 45S5 vs. Calcium hydroxide on *Enterococcus faecalis*. *Iranian Endod J*. 2011;6(1): 29-32.
32. McHugh CP, Zhang P, Michalek S, Eleazer PD. pH required to kill *Enterococcus faecalis* in vitro. *J Endod*. 2004;30(4): 218-219.
33. Attia DA, Farag AM, Afifi IK, Darrag AM. Antimicrobial effect of different intracanal medications on various microorganisms. *Tanta Dent J*. 2015;12(1): 41-47.
34. Hulsmann M, Heckendorff M, Lennon A. Chelating agents in root canal treatment: Mode of action and indications for their use. *Inter Endod J*. 2003;36(12): 810-830.
35. Happsalo M, Shen YA. Current therapeutic options for endodontic biofilms. *Endod Topics*. 2010;22(1): 79-98.
36. Abbaszadegan A, Khayat A, Motamedifar M. Comparison of antimicrobial efficacy of IKI and NaOCL irrigants in infected root canals: An in vitro study. *Iranian Endod J*. 2010;5(3): 101-103.
37. Molander A, Reit C, Dahlen G. The antimicrobial effect of calcium hydroxide in root canals pretreated with 5 % iodine potassium iodide. *Dent Traumatol*. 1999;15(5): 205-209. <https://doi.org/10.1111/j.1600-9657.1999.tb00775.x>
38. Nakajo K, Iwami Y, Komori R, Ishikawa S, Ueno T, Suzuki Y, Takahashi N. The resistance to acidic and alkaline environment of endodontic pathogen *Enterococcus faecalis*. *International Congress Series*. 2005;1284: 191-192.

دراسة معملية بخصوص الفعالية المضادة للميكروبات لهيدروكسيد الكالسيوم وتوليفته مع هيبوكلوريت الصوديوم ضد المكورات المعوية البرازية والمبيضات المبيضة

محمد عيسى¹، نجوى المرغني²

¹قسم العلاج التحفظي وعلاج الجذور ، كلية طب الأسنان ، جامعة طرابلس ، طرابلس ، ليبيا
²قسم المختبرات ، مستشفى طرابلس الجامعي ، طرابلس ، ليبيا

المستخلص

كان الهدف من هذه الدراسة المعملية هو تقييم ومقارنة التأثير المضاد للميكروبات لهيدروكسيد الكالسيوم وتوليفته مع هيبوكلوريت الصوديوم 5% كأدوية داخل القناة ضد المكورات المعوية البرازية و المبيضات المبيضة بعد يوم وثلاثة وسبعة أيام . تم استخدام التلقيح من هذه الكائنات الحية (المكورات المعوية البرازية و المبيضات المبيضة) في زراعة العشب في مولر- هينتون باستخدام أجار دم الأغنام منزوع الرجفان بنسبة 5% وأجار مولر- هينتون. تم تحضير 60 بئراً بعمق 4 ملم للمكورات المعوية البرازية . ثم تم ملء 20 بئراً بهيدروكسيد الكالسيوم، و 20 بئراً مملوءة بمزيج من هيدروكسيد الكالسيوم و 5% هيبوكلوريت الصوديوم ، وتم ملء آخر 20 بئراً بالماء المقطر كعنصر تحكم. تم إجراء إجراء مماثل للمبيضات المبيضة. تم قياس وتسجيل منطقة التثبيط لكل مادة مستخدمة ضد كائن حي معين بعد 1، 3، و 7 أيام . يعتبر هيدروكسيد الكالسيوم مع 5% هيبوكلوريت الصوديوم أكثر فعالية من هيدروكسيد الكالسيوم وحده ضد المكورات المعوية البرازية و المبيضات المبيضة في فترات زمنية مختلفة. كانت المكورات المعوية البرازية أكثر مقاومة من المبيضات المبيضة للأدوية المستخدمة داخل القناة. يعتمد التأثير المضاد للميكروبات على المدة التي تبقى فيها داخل قناة الجذر. كان لخلط هيدروكسيد الكالسيوم مع 5% هيبوكلوريت الصوديوم تأثير مضاد للميكروبات على كل من المكورات المعوية البرازية و المبيضات المبيضة وكان أكثر فعالية ضد المبيضات المبيضة .

الكلمات الدالة: هيدروكسيد الكالسيوم، هيبوكلوريت الصوديوم، المكورات المعوية البرازية، المبيضات المبيضة.